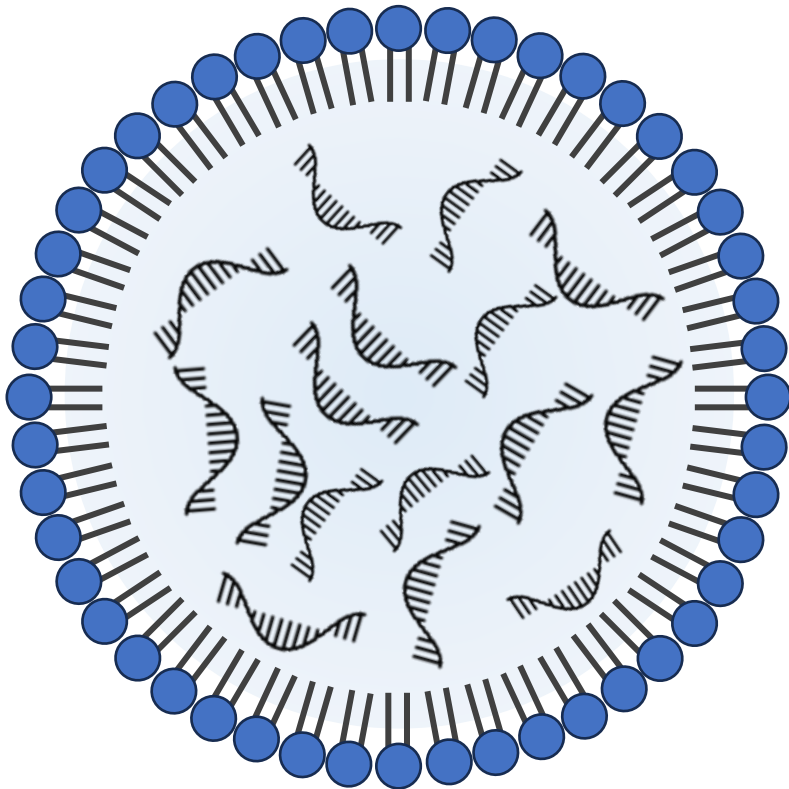


# What are lipid nanoparticles ? (LNP)

## LNP (Lipid Nano Particle) 1※

= LNPs are nanocarriers comprised of lipids that safely deliver pharmaceutical ingredients to target cells.



Hydrophilic head

Hydrophobic Tail



Lipid



Nucleic acid(e.g., mRNA、siRNA)  
or small-molecule

The lipid layer encapsulates the nucleic acid or small molecule

### Usage

- ✓ mRNA vaccines
- ✓ Gene therapy
- ✓ Anti-cancer treatments

Etc.

Particle diameter: 50~100nm

1 – J. Pharm. Sci. Technol. Jpn., 82(2), 63-70 (2022).

# Mechanism of LNP formation

**Lipid** Solution



**RNA**

Aqueous Solution



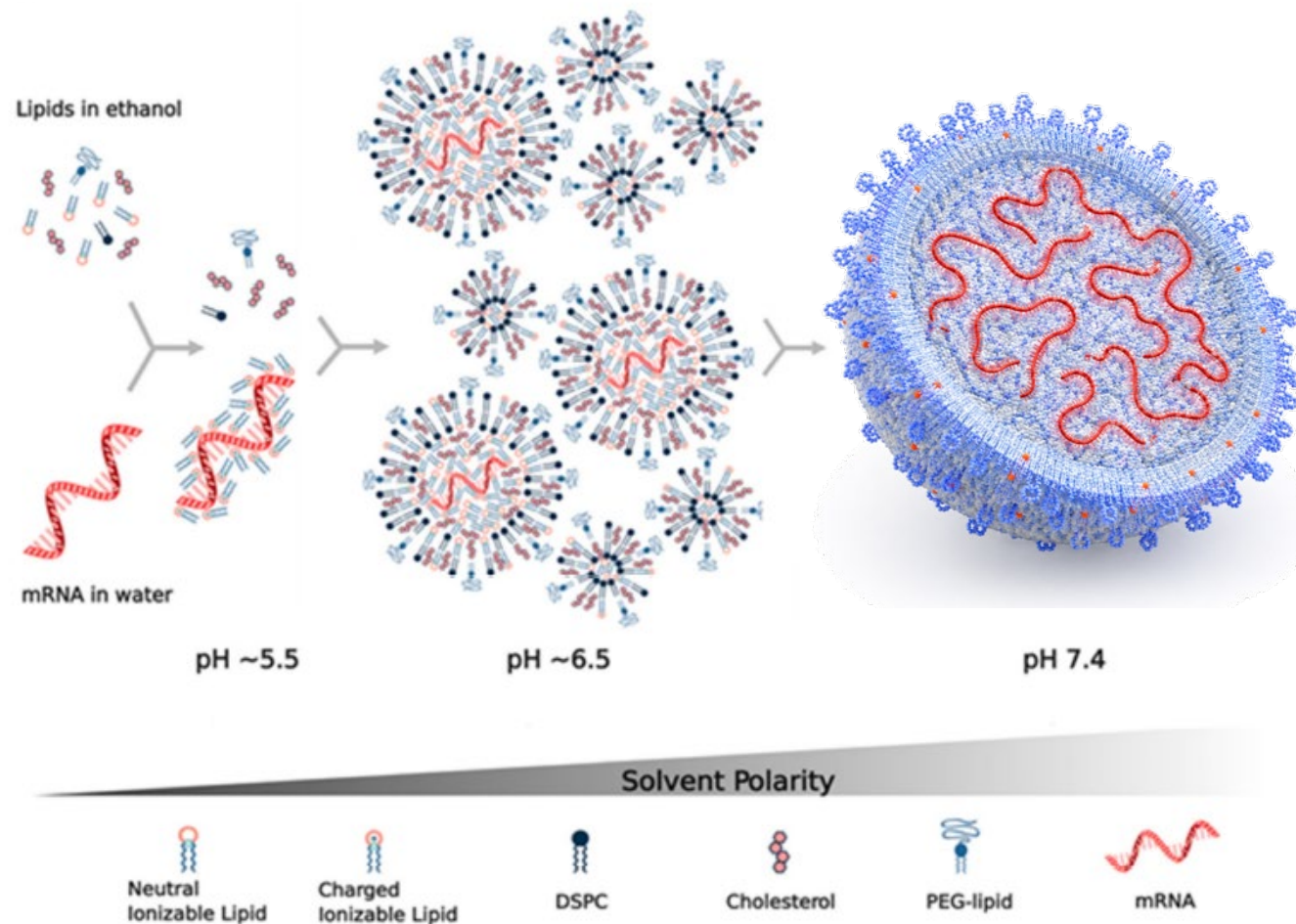
**Rapid Mixing** changes the solvent polarity, causing lipids to self-assemble.



Under acidic conditions, ionized lipids interact electrostatically with mRNA.

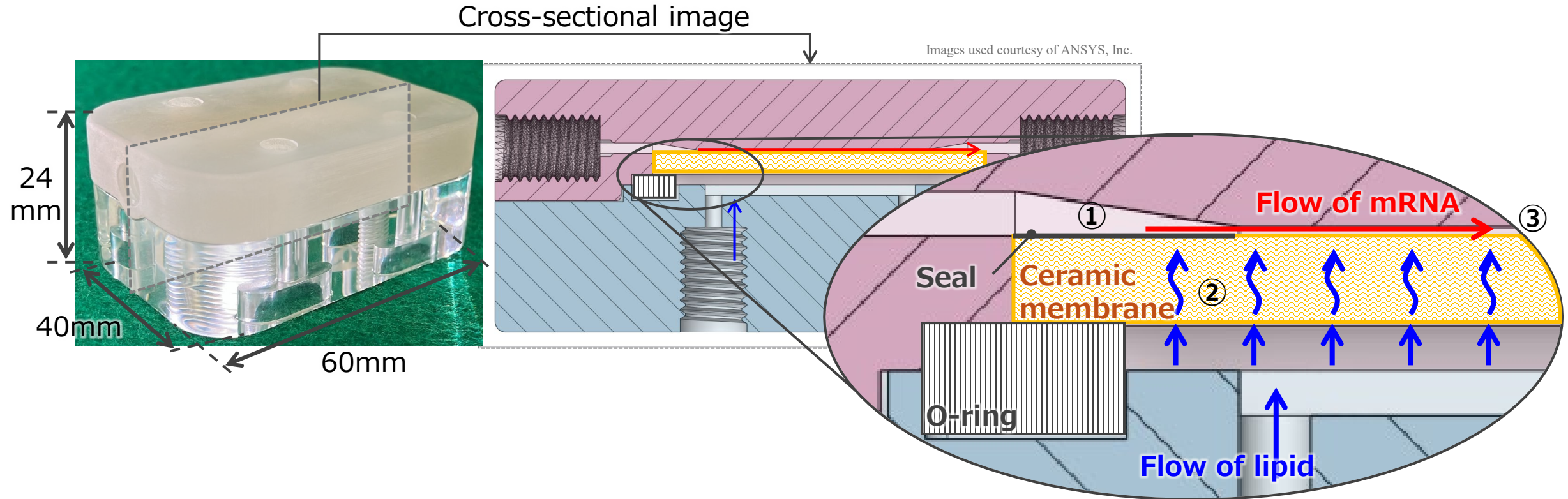


At neutral pH, the particle surface stabilizes, forming mRNA-containing LNPs.



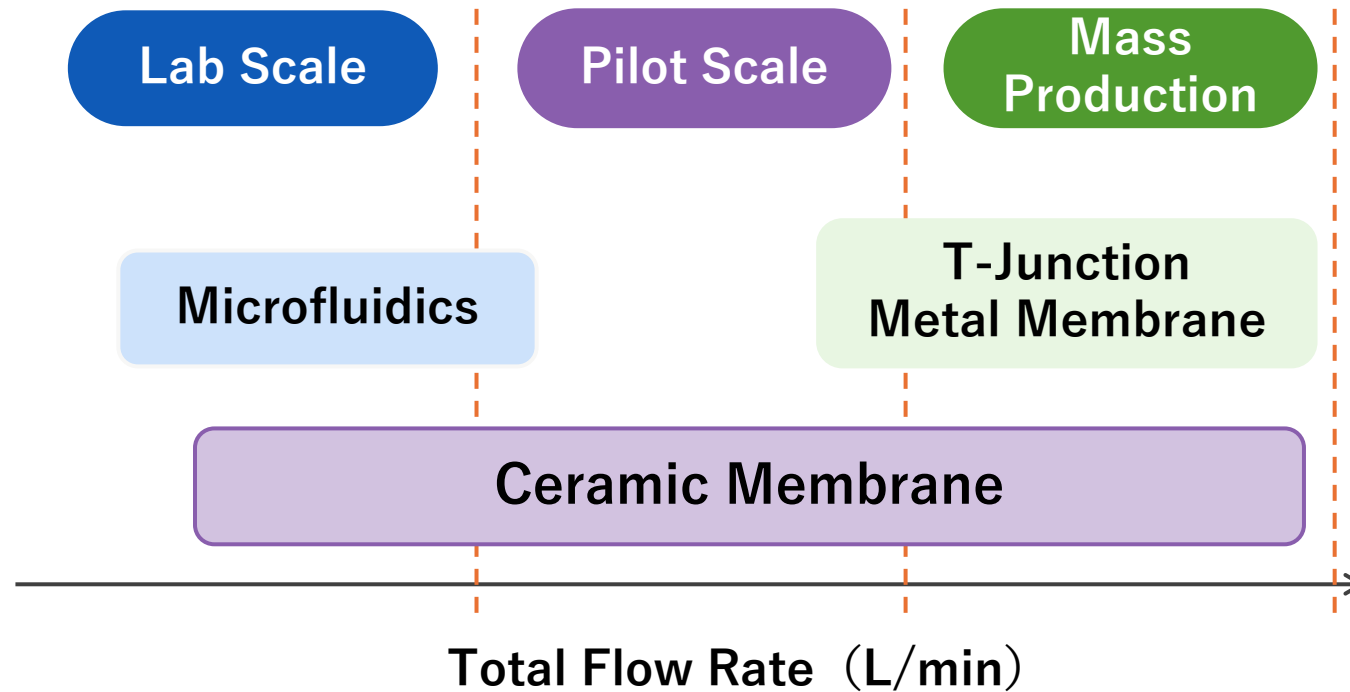
Buschmann, Michael D., et al. "Nanomaterial delivery systems for mRNA vaccines." Vaccines 9.1 (2021): 65. より引用・一部改変

# NGK's Ceramic Membrane-Based LNP Manufacturing Device



- ① The API solution(i.e. mRNA solution) flows across the surface of the ceramic membrane from the left side of the device.
- ② The lipid solution is fed through the bottom of the device and permeates through the ceramic membrane towards the top.
- ③ As the lipid solution permeates through the numerous pores of the membrane it mixes with API solution flowing across the membrane surface leading to LNP formation. The LNPs are collected from the outlet on the right side of the device.

# Scalability: Issues with current methods and their solution



**Microfluidics** : Difficult to scale up due to LNP aggregation making high flow rates difficult

**Metal Membrane** : Difficult to scale down as limitations in pore diameters makes forming LNPs at low flow rates difficult.

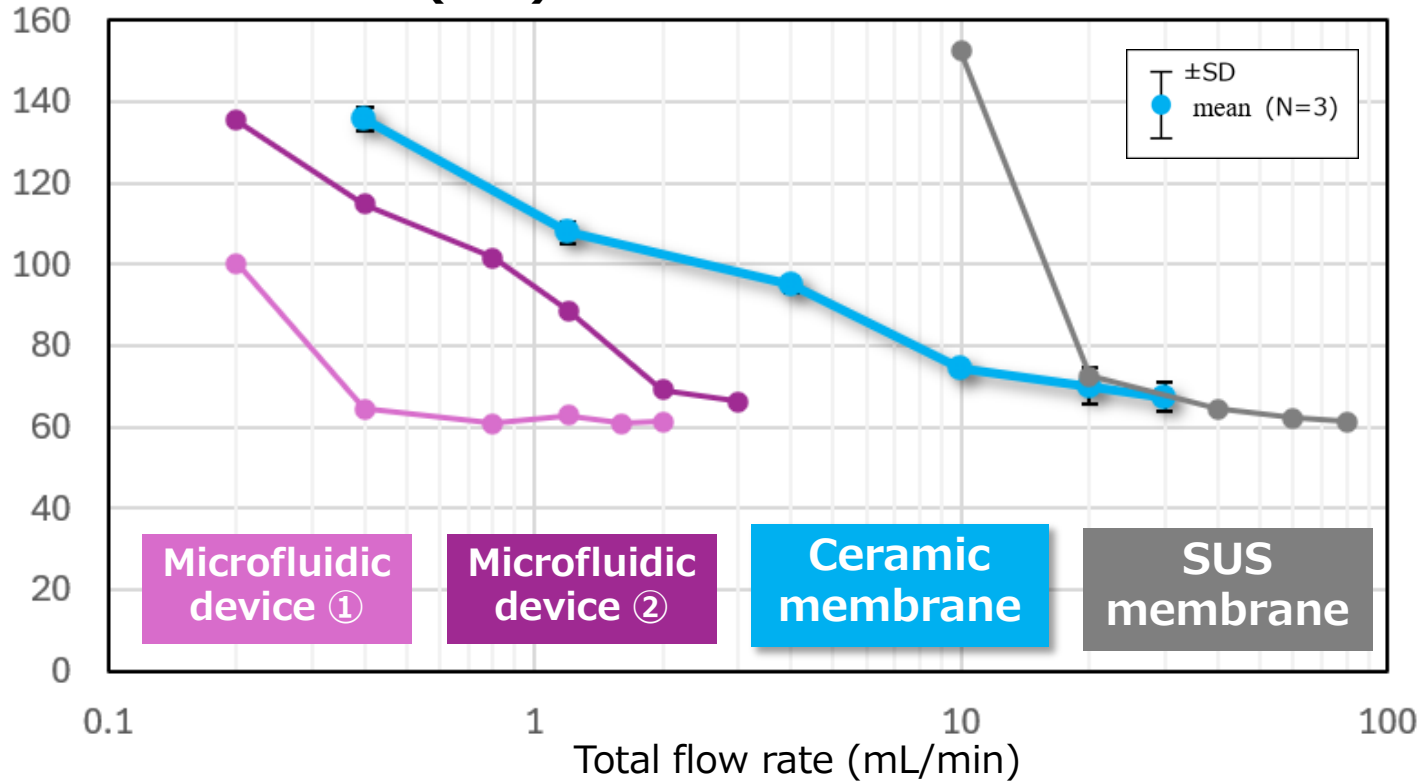
**T-Junction**: Difficult to scale down as LNPs are formed by rapid mixing at high speeds makes forming LNPs at low flow rates difficult.

**Ceramic Membrane** : Adjusting membrane area and porosity enables scalability from laboratory development to commercial production.

# The solution to the issue of scalability

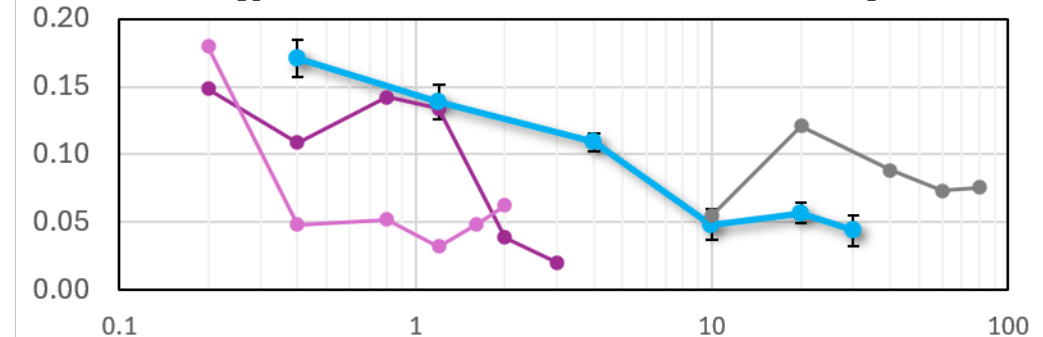
Evaluated by TAIYO HOLDINGS CO., LTD.

## Particle size(nm)

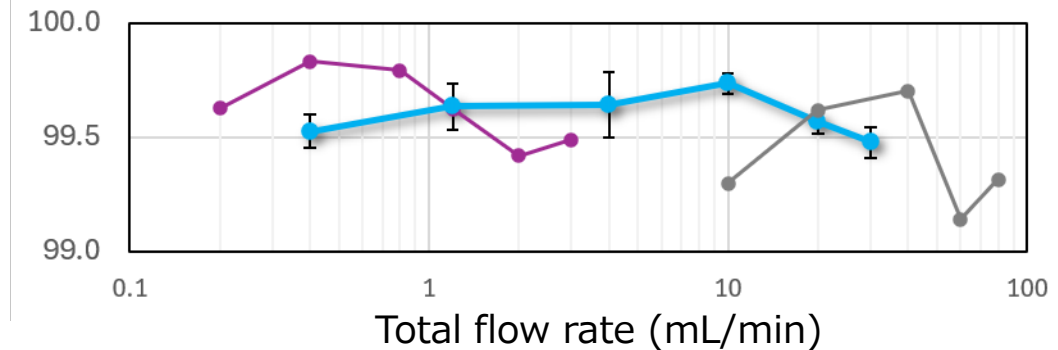


Microfluidic device ① : iLiNP-1.0S  
Microfluidic device ② : iLiNP-1.0SW  
SUS membrane : AXF-mini

## PDI (particle size distribution)



## Encapsulation efficiency(%)

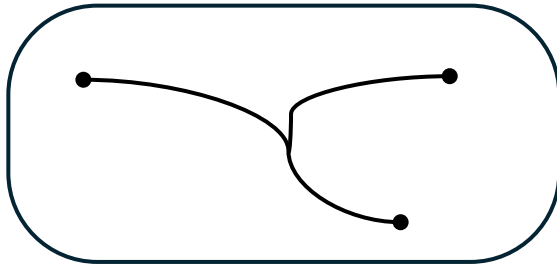


**Ceramic membrane technology maintains particle characteristics across a broad operating flow-rate range.**

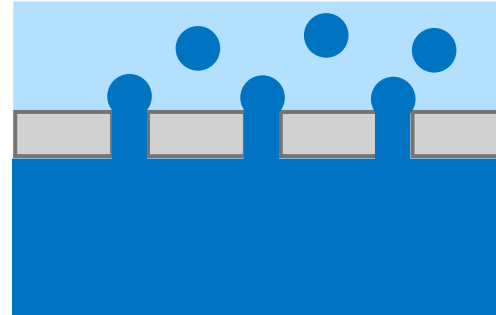
In this evaluation, a Spikevax-type lipid formulation and polyA as an mRNA substitute was used.

# Single-Use Operation, Chemical resistance: Issues with current methods and their solution

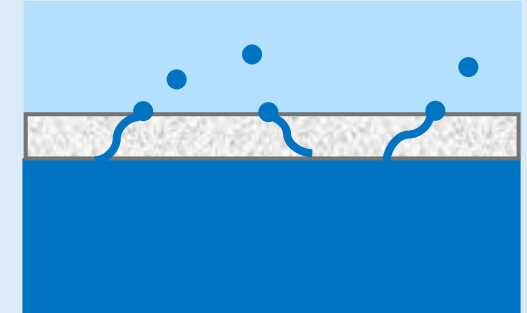
Microfluidics



Membrane Emulsification  
~Metallic membrane~



Membrane Emulsification  
~Ceramic membrane~



Single-Use  
Operation

Single usage considering  
the cost and replicability of  
microchip

Single-use operation is  
challenging due to  
membrane cost.

Replaceable membrane  
design enables single-use  
operation.

Chemical  
Resistance

If the material is plastic  
certain solvents may cause  
channel deformation,  
altering mixing conditions.

High chemical stability  
due to the material being  
metal.

High chemical stability due  
to the material being  
ceramic.

# In vitro evaluation between conventional method and NGKs method

Evaluated at the Faculty of  
Pharmaceutical Sciences,  
Kyushu University,



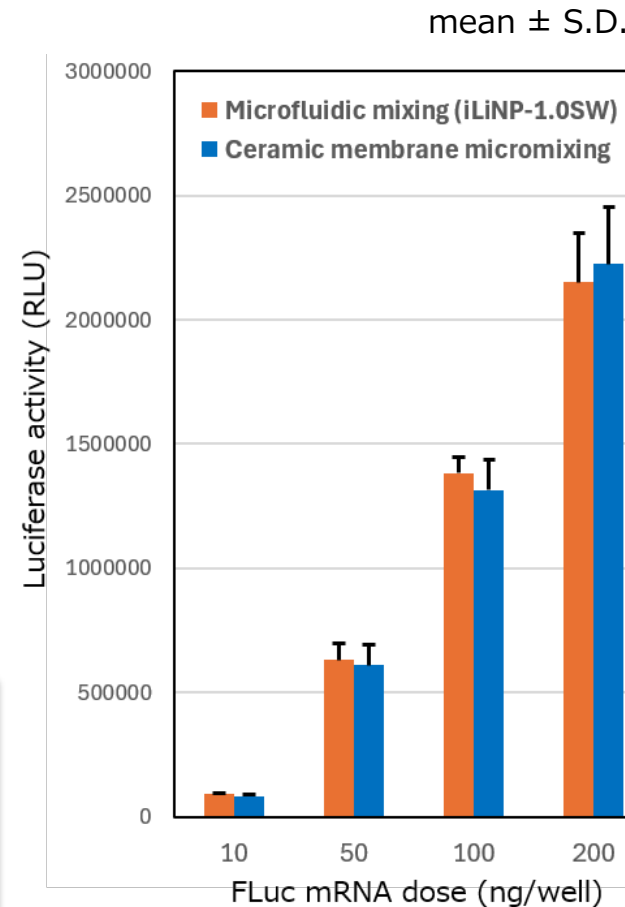
| LNP Characteristics      |    | Microfluidic<br>(iLiNP-1.0SW) | Ceramic Membrane |
|--------------------------|----|-------------------------------|------------------|
| Particle size            | nm | 84.5                          | 84.1             |
| PDI                      | —  | 0.131                         | 0.077            |
| Zeta potential           | mV | +2.8                          | +2.5             |
| Encapsulation efficiency | %  | 95.0                          | 95.9             |

Concentration : 10 $\mu$ g FLuc-mRNA/mL  
 Dosage : 1, 5, 10, 20 $\mu$ L/well  
 No. of cells : 2 $\times$ 10<sup>4</sup> cells/50 $\mu$ L/well  
 Measurement : 24hrs  
 N : n=5

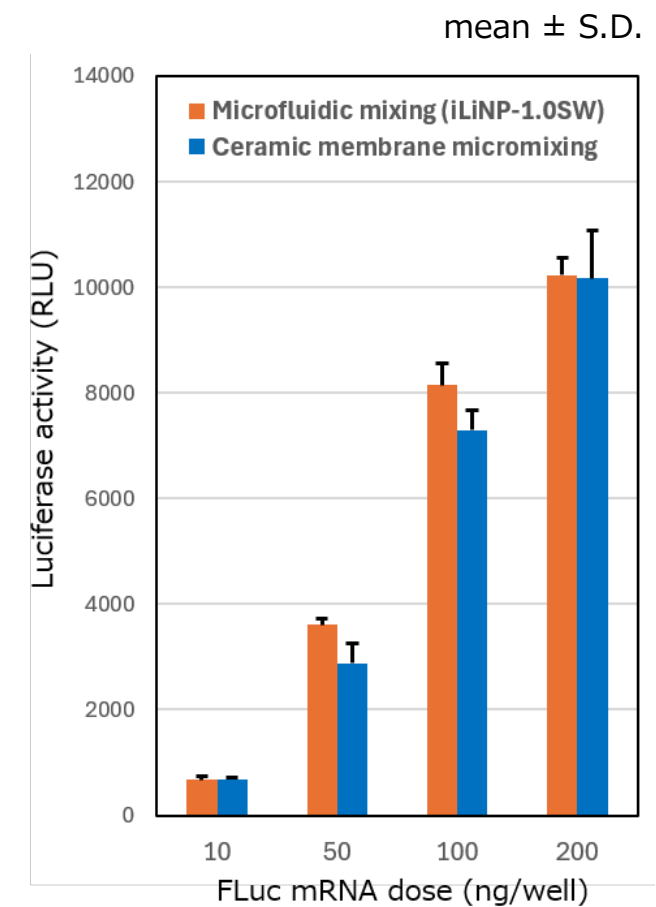


**The LNPs prepared using the ceramic membrane emulsification method showed *in-vitro* FLuc expression comparable to conventional microfluidic systems**

## RAW264.7 (Mouse macrophage-like cells)



## THP-1 (Human monocytic cells)

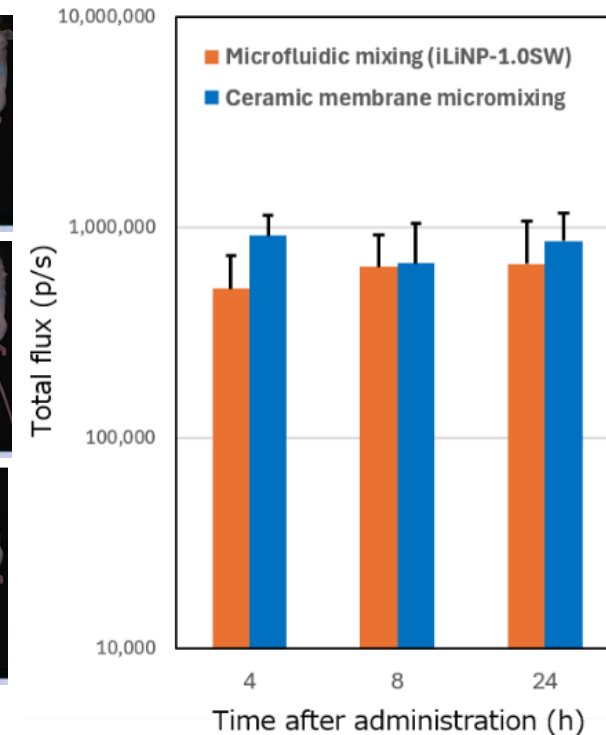
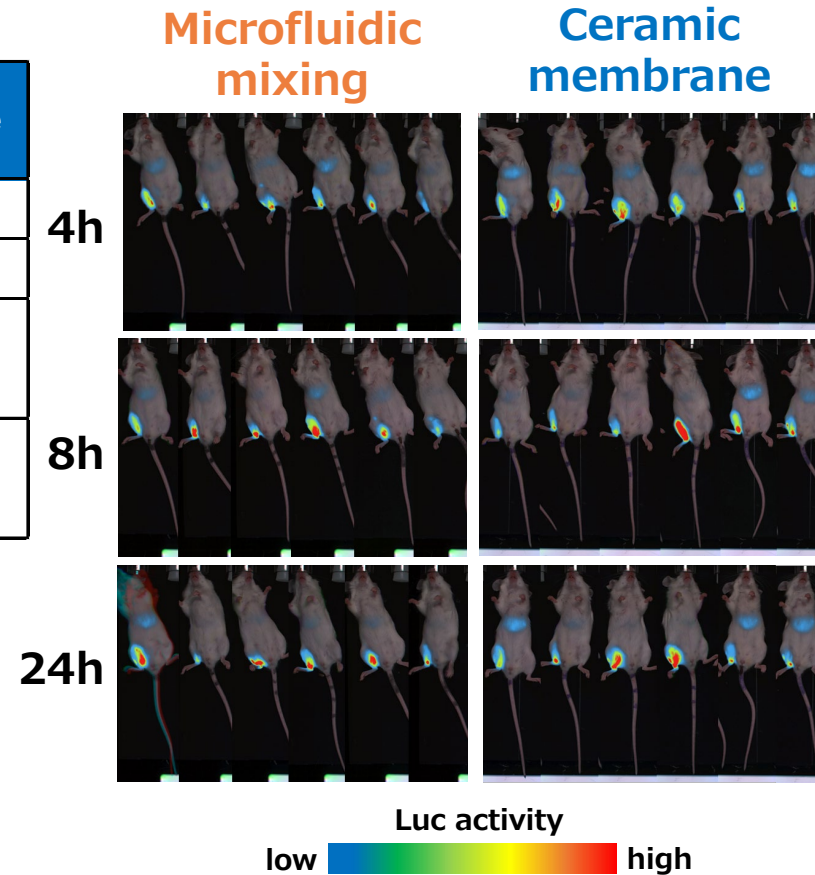


# In vivo evaluation: FLuc expression activity

Evaluated at the Faculty of  
Pharmaceutical Sciences,  
Kyushu University,

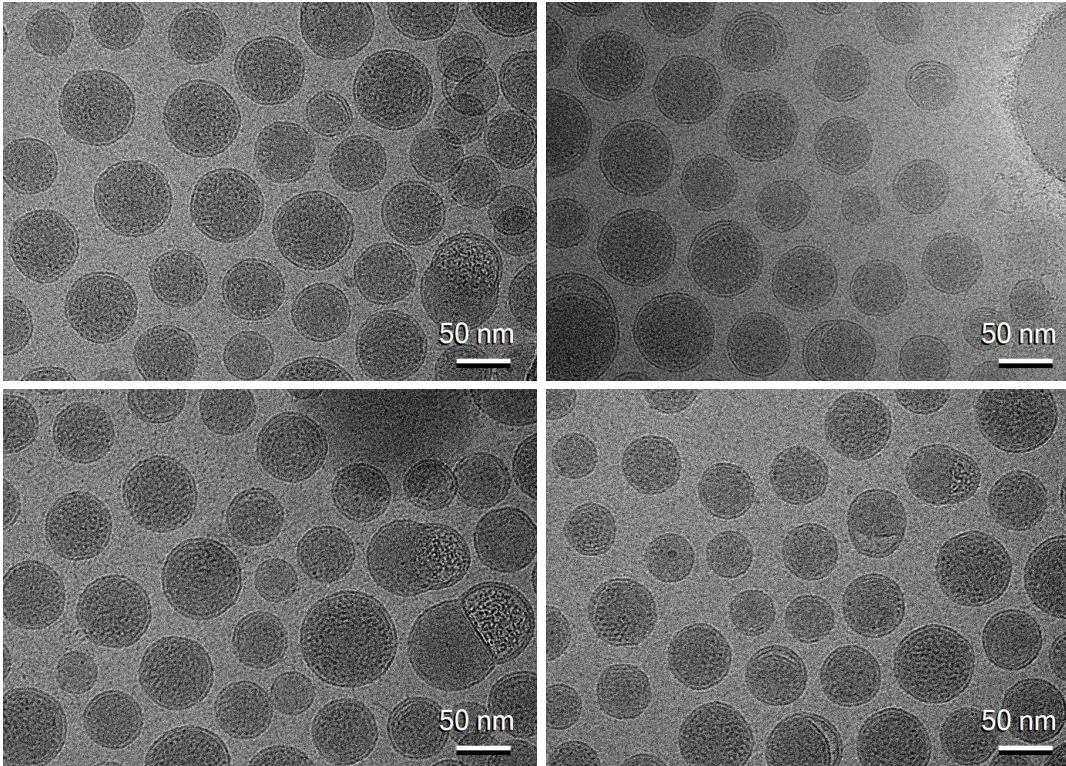
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| PDI                      | —  | 0.131                         | 0.077            |
| Zeta potential           | mV | +2.8                          | +2.5             |
| Encapsulation efficiency | %  | 95.0                          | 95.9             |

LNP conc. : 10 $\mu$ g FLuc-mRNA/mL  
Mouse : BALB/c Female, 6 weeks of age  
Dosage : 0.1 $\mu$ g mRNA/10 $\mu$ L/mouse  
Injection site : calf muscle  
Measurement : 4h, 8h, 24h  
n : n=6

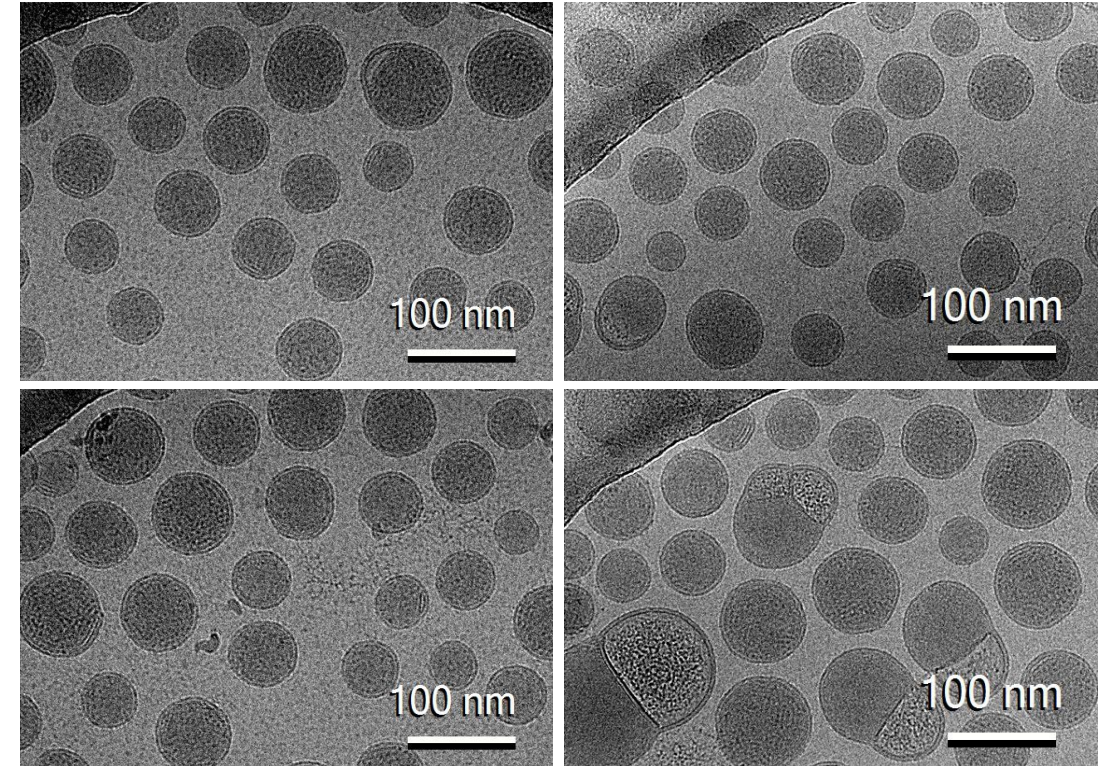


The LNPs prepared using the ceramic membrane emulsification method showed *in-vivo* FLuc expression activity comparable to conventional microfluidic systems.

**Microfluidic** (iLiNP-1.0SW)



**Ceramic Membrane**



**LNPs prepared using the ceramic membrane exhibited structural characteristics comparable to those produced using conventional microfluidic systems.**